


# T lymphocytes and preeclampsia: The potential role of T-cell subsets and related MicroRNAs in the pathogenesis of preeclampsia

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## Funding information

This work was supported by Molecular Medicine Research Center, Biomedicine Institute, Tabriz University of Medical Sciences, Tabriz, Iran (Grant No. 59958).

## Abstract

Innate and adaptive immune systems have a crucial role in initiating and progressing some pregnancy disorders such as preeclampsia (PE), which is one of the pregnancy-specific disorders that could result in neonatal and maternal morbidity and mortality. The dysregulation of the spiral artery and inadequate trophoblast invasion lead to PE symptoms through producing various inflammatory cytokines and anti-angiogenic factors from the placenta. T lymphocytes play a special role in the epithelium and stroma of the human endometrium. CD4+ T helper (Th) cells, Th1/Th2, and Th17/Treg balance mainly contribute to the establishment of a pregnancy-favorable environment. This review examined the dysregulation of some cytokines produced from T cells, the dysregulation of the transcription factors of Th cells, the expression of chemokine receptors on T cells, as well as the effects of some factors including vitamin D on the activity of T cells, and finally, the dysregulation of various miRNAs related to T cells, which could cause PE.

## KEYWORDS

miRNA, preeclampsia, pregnancy, T cell

## 1 | INTRODUCTION

Adaptive and innate immune systems have a major role in the initiation and development of different types of pregnancy disorders such as preeclampsia (PE).<sup>1</sup> Approximately, 5%-8% of pregnant people suffer from PE worldwide. PE is one of the pregnancy-specific disorders which could result in neonatal and maternal morbidity and mortality.<sup>2</sup> It is diagnosed by different symptoms such as thrombocytopenia, pulmonary edema, renal insufficiency, proteinuria, visual or cerebral symptoms, and the presence of de novo hypertension after the 20th gestational week.<sup>3,4</sup> PE can be categorized into early- (before 34 weeks) and late-onset (after 34 weeks) types. The dysregulation of the spiral artery and inadequate trophoblast invasion lead to the symptoms of PE by producing various inflammatory cytokines and anti-angiogenic factors from the placenta.<sup>5</sup> In recent years, research has expanded on the disorders of different subtypes of T lymphocyte cells, the dysregulation of releasing cytokines from T cells, and the expression of microRNAs (miRNAs) related to T cells in PE. This review also focuses on the advances in our knowledge about the role of T lymphocytes in the pathogenesis of PE.

## 2 | LYMPHOCYTES AND PREGNANCY

Lymphocytes are the largest resident immune cell population found in the decidualized endometrium and include uterine or decidual natural killer (NK) cells and the T-cell subsets, namely CD4 T helper 1 (TH1), Th2 CD8, T regulatory (Treg), and Th17 cells. T lymphocytes are located in the decidual stroma and glandular epithelium where they play a critical role in the establishment of a favorable pregnancy environment.<sup>6</sup> Th1/Th2 and Th17/Treg cell balance have an essential part in the establishment of a pregnancy-favorable environment. During the peri-implantation period, a controlled shift toward Th1 responses is involved in immune surveillance and avoids the excessive invasion of trophoblast.<sup>7,8</sup> After the placental implantation, making a shift toward Th2 is essential for the preservation and development of a normal embryo, the suppression of Th17 and Th1 cells by releasing interleukin (IL)-13 and IL-4, respectively, and an increase in allograft tolerance.<sup>8,9</sup> Additionally, Treg cells are the main factors for immunological tolerance, the acceptance of the fetus by the mother's immune system, and embryo implantation. Peripheral and decidual Treg cells increase during normal pregnancy.<sup>10,11</sup> Tregs exert suppressive functions through different mechanisms, including the modulation of antigen presentation, the cytolysis of target cells, and the secretion of inhibiting cytokines.<sup>12,13</sup> Treg cells prevent the operation and proliferation of Th17 and Th1 cells via different mechanisms, including programmed cell death protein 1 (PD-1) and the Ca<sup>+</sup> influx-clapin-caspase 1 pathway. Over the years, studies have shown that different subsets of Tregs have various effects on pregnancy. Zenclussen et al revealed the exact type of thymus Treg (tTreg) cells that are vital for the establishment of pregnancy.<sup>14</sup> In this regard, the activity of induced Treg (iTregs) is demonstrated in

the late fertility stages.<sup>15</sup> During pregnancy, Th17 cells make immunity against extracellular pathogens and induce NK cell activation.<sup>8</sup> IL-17 promotes tissue invasion and progesterone secretion.<sup>16</sup> Low levels of Th17 cells are observed during normal pregnancy as compared to infertile women, and Th17 cell levels do not vary in fertility processes. In addition, the level of Th17 in the decidual is higher compared to the peripheral blood.<sup>17,18</sup> According to a study, the placental trophoblast through making a shift toward Th2 responses and inhibition of Th17 cells leads to the maintenance of pregnancy.<sup>19</sup>

## 3 | Th1 AND Th2 CELL FUNCTION AND RELATED FACTORS IN PE

Preeclampsia patients exhibit chronic inflammation.<sup>20</sup> The balance between the functions of Th1/Th2 changes toward a Th1-dominated phenotype, and a high level of IL-12 can cause the severity of PE.<sup>21</sup> In this respect, the ratio of Th1/Th2 cells elevates as well.<sup>22</sup> Th1 cells have a critical role in systematic inflammation by secreting the amounts of tumor necrosis factor-alpha (TNF- $\alpha$ ), interferon-gamma (IFN- $\gamma$ ), IL-1 $\beta$ , and IL-12. The domination of Th1 responses can induce chronic inflammatory reactions in the fetal-maternal interface, endothelial dysfunction, and the impairment of placentation.<sup>23,24</sup> Some studies represented the transfusion of Th1 cells into normal pregnant mice that demonstrated PE symptoms including proteinuria, blood pressure, inflammation of the decidua, and changes in kidney functions. However, the transfer of Th1 cells into non-pregnant mice does not induce any alterations in renal function and blood pressure.<sup>22</sup>

T-bet and GATA binding protein 3 (GATA-3) as the transcription factors of Th1 and Th2 cells regulate the expression of Th1 and Th2 cytokine, respectively, and have critical roles in the differentiation of Th cells.<sup>25</sup> According to a study, higher and lower percentages of T-bet and GATA-3 were found in peripheral and decidual T cells in PE patients, respectively, as compared to normal pregnant.<sup>25,26</sup>

## 4 | Treg AND Th17 CELL FUNCTION AND RELATED FACTORS IN PE

It has been shown that number of Treg cells is decreased and their function impaired in peripheral blood and decidua in PE patients.<sup>27,28</sup> There is an altered prevalence of Treg subtypes in PE compared with healthy pregnant. The prevalence of fully functional effector Tregs (CD4<sup>+</sup> FoxP3hi CD45RA<sup>-</sup>) is reduced, while naïve Tregs (CD4<sup>+</sup> FoxP3<sup>+</sup> CD45RA<sup>+</sup>) remain to be unaffected.<sup>29</sup> Feger et al<sup>30</sup> described HLA-G<sup>+</sup> Treg cell subsets. These cells do not express Foxp3 and CD25 molecules, are of hypo-proliferative and thymic-derived types, and are detected in HIV-1 infection, multiple sclerosis, and transplantation.<sup>31</sup> The frequency of CD4<sup>+</sup> HLA-G<sup>+</sup>, CD8<sup>+</sup> HLA-G<sup>+</sup>, and CD4<sup>+</sup> CD25<sup>+</sup> CD127<sup>low</sup> peripheral cells in the PE group is lower compared to a healthy pregnant group.<sup>32</sup> According to these results,

decreased frequencies of regulatory T cells can lead to immunological maladaptation and inadequate tolerance to the fetus in PE.<sup>33</sup> Higher numbers of CD4<sup>+</sup> CD25<sup>high</sup> FoxP3<sup>+</sup> CD279<sup>+</sup> identified as exhausted Tregs were found in peripheral T cells in PE patients.<sup>34,35</sup> The exhausted T cell is a state of T-cell dysfunction that decreases the strength of the immune system in controlling infection and cancer.<sup>36</sup> CD279 (PD-1) expression has been considered as a marker of T-cell exhaustion.<sup>37</sup> PD-1 expression on T cells is associated with restricted proliferative capacity and decreased suppression of the immune system.<sup>29</sup> Higher expression of PD-1 and interactions with its ligand (PDL-1) generate a strong co-inhibitory signal in exhausted T cells.<sup>36</sup> Thereby, this molecule may involve in reducing the number and function of Tregs in PE.<sup>34</sup> During pregnancy, the PD-1/PDL-1 pathway plays a major role in maintaining feto-maternal tolerance by regulating the immune response.<sup>38</sup> According to Zhang et al, the PD-1/PDL-1 pathway contributes to the Treg/Th17 balance during pregnancy by promoting Treg cell generation and preventing Th17 proliferation. However, PD-1/PDL-1 pathway dysfunction may exert a role in the Treg/Th17 imbalance in peripheral blood and decidua, leading to the development of PE.<sup>39</sup> The Treg-related factors are summarized in Table 1.

Th17 cells are involved in pro-inflammatory and autoimmune diseases by secreting IL-17 cytokine.<sup>40</sup> Studies have confirmed that Th17 cell populations and their cytokines are elevated in PE women.<sup>41</sup> Moreover, both TH17 and IL-17 increase in reduced uterine perfusion pressure (RUPP) rats.<sup>42</sup> The dysregulation in NK cell subtypes, agonistic autoantibodies to the ANGII type 1 receptor (AT1-AA), inflammatory cytokines, and placental oxidative stress

are the main factors for proteinuria, high blood pressure, and low birthweight in PE patients.<sup>43</sup> As reported by a study, separating and transferring RUPP Th17 to normal pregnant (NP) rats can induce the levels of inflammatory cytokines such as IL-17, IL-6, IFN- $\gamma$ , TNF- $\alpha$ , cytolytic NK functions, markers of blood pressure, placenta perforin, and AT1-AA production, and placenta oxidative stress including reactive oxygen species.<sup>44</sup> Moreover, Travis et al investigated the role of IL-17, which is secreted by Th17, in hypertension and pregnant rats. They concluded that IL-17 infusion into a subset of NP rats can induce the activation of NK cells, the plasma level of TNF- $\alpha$ , the reduction of placental vascular endothelial growth factor and fetal weight, the production of a high level of granzymes A and B, and finally, the impairment of the vascular reactivity of uterine arteries and PE defects.<sup>45</sup> The Th17-related factors are presented in Table 1.

Orphan nuclear receptor  $\gamma$ t (ROR $\gamma$ t) and forkhead box p3 (Foxp3) are specific transcription factors that are involved in the production of Th17 and Treg cells.<sup>46</sup> Eghbal-fard et al<sup>46</sup> approved an increasing Th17/Treg ratio in PE by detecting the percentages of CD4<sup>+</sup> CD25<sup>+</sup> CD127<sup>-</sup> and CD4<sup>+</sup> IL-17<sup>+</sup> cells. Additionally, the mRNA level of ROR $\gamma$ t increased while that of Foxp3 decreased in peripheral and decidual T cells in the PE patients as compared to healthy pregnant.<sup>25,46</sup> An imbalance between Treg and Th17 cells can induce chronic inflammation in PE.<sup>33</sup> Moreover, the mRNA levels and the secretion of inflammatory cytokines in peripheral blood mononuclear cells (eg, IL-6, IL-23, and IL-17) are meaningfully higher in PE women. On the other hand, anti-inflammatory cytokines such as IL-10 and transforming growth factor-beta (TGF- $\beta$ ) are remarkably lower in PE.<sup>46</sup>

**TABLE 1** Dysregulation of T-cell subsets and related factors in PE

Dysregulation of various factors in PE	Function
Th1	Inducing chronic inflammatory reactions in the fetal-maternal interface by secreting the amounts of TNF- $\alpha$ , IFN- $\gamma$ , IL-1 $\beta$ , and IL-12
T-bet, GATA-3	High expression of T-bet and low expression of GATA-3 may lead to Th1/Th2 imbalances toward to Th1
PD-1/PD-L1	PD-1/PD-L1 pathway dysfunction may exert a role in the Treg/Th17 imbalance in peripheral blood and decidua
IL-17	IL-17 is found by Th17 and elevation in angiotensin II type I receptor (AT1-AA) production
Th17	Inducing cytolytic NK cell's function, placenta perforin production, IFN- $\gamma$ , and TNF- $\alpha$ cytokines
ROR $\gamma$ t, Foxp3	High expression of ROR $\gamma$ t and low expression of Foxp3 may lead to Th17/Treg imbalances toward to Th17
iNKT	Have an augmented cytotoxic potential along with exhibiting Th1 dominant profile
CXCL16	Interaction with CXCR6 receptors on T cells and subsequently inflammation
IFN- $\gamma$	Inducing the expression of CXCL9 chemokines which are involved in inflammatory responses
25 (OH) vitD	Increasing IL-6 and decreasing TGF- $\beta$ secretion from Th1 and Treg
Adenosine	Th1/Th2 balances toward Th1 and subsequently hypertension
DCR3	Suppressing Th2 cells function and Th1/Th2 imbalances

## 5 | NKT CELLS IN PE

These cells are a subset of lymphocytes that express NK cell receptors and can be classified into the invariant NKT (iNKT) cells with the T-cell receptor (TCR) alpha-chain (TCR $\alpha$ ) or variant NKT (vNKT) cells without the expression of the invariant T-cell receptor (TCR) alpha-chain.<sup>1</sup> NKT cells can control the immune response in decidua and peripheral blood through the production of Th1-type cytokines (IFN- $\gamma$ ) and Th2-type cytokines (IL-4).<sup>24</sup> In the decidua, NKT cells can regulate Th1/Th2 balance and are involved in maintaining pregnancy.<sup>1</sup> An alteration in the balance of NK cell activating and inhibitory receptors on peripheral iNKT cells in women with PE indicates a lower chance of inhibitory signal transduction as compared with healthy pregnancies.<sup>47</sup> Thereby, the frequency of activated peripheral iNKT cells is fully higher in PE women, and they have an augmented cytotoxic potential along with exhibiting Th1 dominant profile, suggesting the important role of iNKT cells in making a systemic inflammation.<sup>24,48</sup>

## 6 | CHEMOKINES AND PE

Chemokines are a superfamily of chemotactic cytokines that have an important impact on the regulation of leukocyte and lymphocyte transport from the bone into the inflammation sites. The dysregulation of chemokine expression has been attributed to some diseases such as PE, as well as habitual and spontaneous abortions.<sup>49</sup> CXC chemokine ligand (CXCL) 16 is a plasma membrane chemokine and consists of a short cytoplasmic tail, a single transmembrane helix, and a chemokine domain with a glycosylated mucin-like stalk.<sup>50</sup> CXCL16 are expressed on smooth muscle cells, endothelial cells, fibroblasts, dendritic cells, B and T cells, macrophages, and platelets.<sup>51</sup> Some of the functions of CXCL16 include being a chemo-attractant for cells carrying chemokine receptor CXCR6, which is known as Bonzo, inducing cell migration into inflammatory areas, and promoting matrix metalloproteinases, which cause the degradation of the matrix.<sup>52</sup> Previous research reported that CXCL16 expression increases in inflammatory diseases, endothelial dysfunction, and hepatic damages.<sup>53</sup> Increased CXCL16 expression has been shown to be associated with inflammation and is induced by inflammatory mediators such as TNF- $\alpha$  and IFN- $\gamma$ . Tok et al<sup>54</sup> showed that the serum level of CXCL16 is meaningfully higher in PE patients, and there exists a positive correlation between CXCL16 and renal and hepatic damages in PE. Elevated CXCL16 levels could be related to systemic inflammation which is involved in the pathogenesis of PE. This study suggested that the CXCL16/CXCR6 axis can be a predicting marker for the development of PE.

Interferon-gamma is a pro-inflammatory cytokine produced by immune cells such as Th1 cells, CD8+ lymphocytes, and NK cells.<sup>55</sup> It is well known that IFN- $\gamma$  is highly expressed in serum/plasma of PE.<sup>56</sup> IFN- $\gamma$  can induce CXCL9 and CXCL10 chemokines, which have a major impact on the recruitment of immune cells into infected/inflamed organs.<sup>57</sup> CXCL9 and CXCL10 exhibit anti-angiogenic features.<sup>58</sup> It has been shown that in the plasma of PE women the levels of CXCL9 and CXCL10 are increased, in line with the increase in

IFN- $\gamma$ .<sup>59</sup> Similarly CXCL9 has been shown to increase in the PE placenta while CXCL10 was unaffected.<sup>60</sup>

Collectively, these findings suggest that the CXC chemokines may contribute to the inflammatory pathogenesis of PE and may present good therapeutic targets.

## 7 | OTHER FACTORS AND PE

### 7.1 | Vitamin D

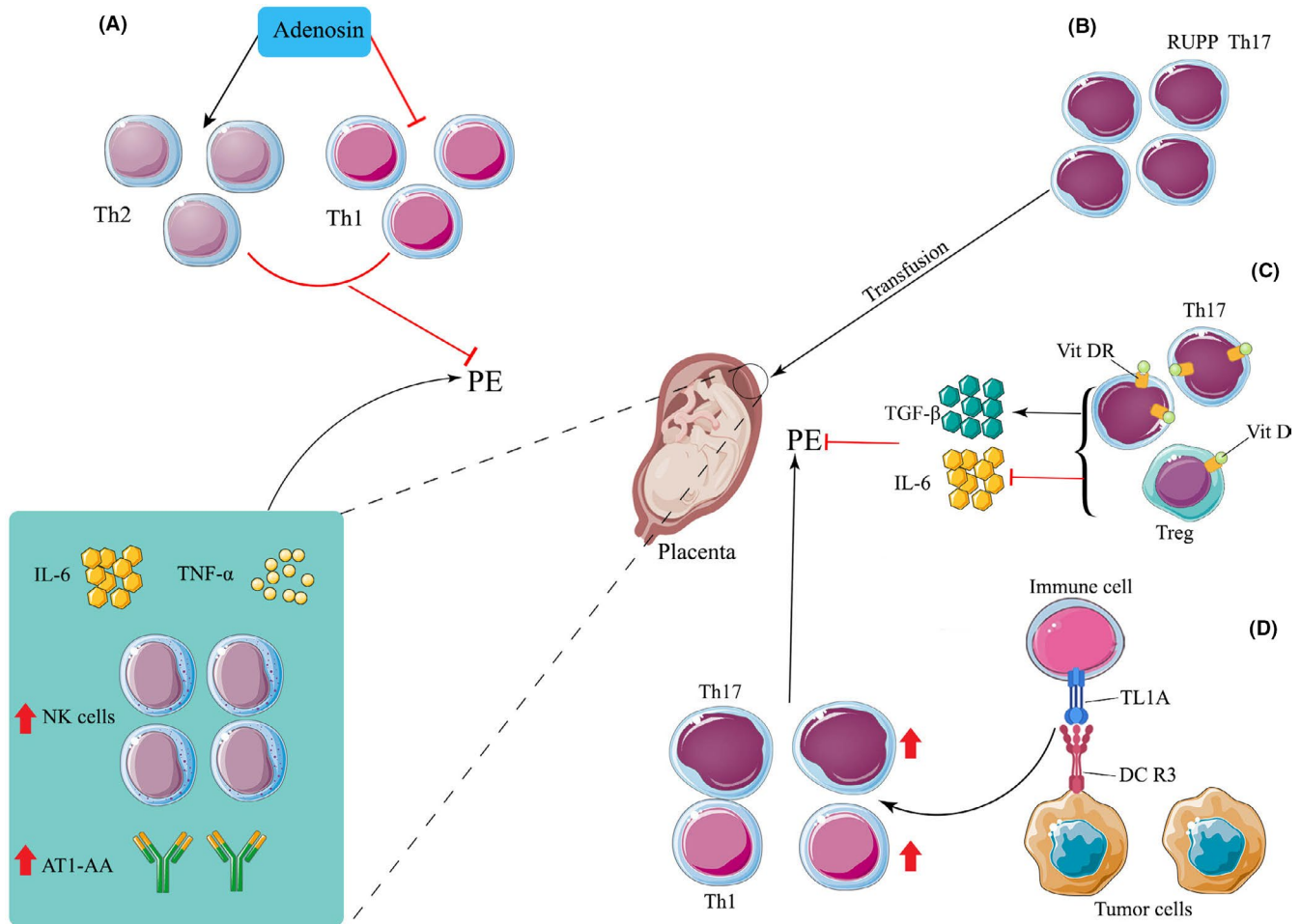
Vitamin D is a critical modulator of vital biological events including immune function, hormone secretion, cell differentiation, and proliferation.<sup>61</sup> It also has a role in the bone synthesis, metabolism, and the regulation of phosphor-calcium metabolism.<sup>62</sup> The vitamin D receptor is expressed on some immune cell members (Th17 and Treg).<sup>63</sup> Vitamin D impedes the lymphocyte proliferation and cytotoxic cell stimulation by alloantigen in the maternal-fetal interface<sup>64</sup> and meaningfully decreases inflammation in PE<sup>65</sup> by the expansion of Treg and Th2 responses.<sup>66</sup> Furthermore, vitamin D inhibits the IL-6 secretion and increases the expression of TGF- $\beta$ .<sup>67,68</sup> As reported by some studies, the level of plasma 25 (OH) D is lower in women with PE as compared with normal pregnancy.<sup>69-71</sup> Moreover, a lower level of vitamin D can be a risk factor for severe PE.<sup>72-74</sup> According to Muyayalo et al<sup>66</sup>, lower levels of vitamin D in PE patients can cause Treg/Th17 cell imbalance and promotion of IL-6 levels.

### 7.2 | Adenosine

Adenosine is produced in reaction to hypoxia and ischemia in the placenta and is a degradative metabolic agent of adenine nucleotides.<sup>75</sup> In addition, it has several roles in different regulatory processes (eg, the regulation of local blood flow and metabolic rate) and has a strong influence on shifting the Th1/Th2 balance toward Th2 dominance and immune-triggered cytokine production.<sup>76</sup> Consequently, adenosine can have a protective role at the maternal-fetal interface. However, plasma adenosine concentrations remarkably increase in PE women<sup>77</sup> and can involve in the regulation of Th1/Th2 ratio imbalance based on the severity of hypertension.<sup>78</sup>

### 7.3 | Decoy receptor 3

Decoy receptor 3 (DCR3; Figure 1) is introduced as tumor necrosis factor receptor superfamily member 6B (TNFRSF6B)/TR6/M68 and a soluble receptor that can neutralize inflammation and apoptosis inducers such as TNF-like molecule 1A (TL1A/TNFSF15), Fas ligand (FasL/CD95L/TNFSF6), and LIGHT (TNFSF14).<sup>79,80</sup> The expression of DCR3 on tumor cells can lead to evasion from immune-cytotoxic attack by preventing the LIGHT-mediated apoptosis and the Fas ligand.<sup>81</sup> According to one study, DCR3 can bind to TL1A on immune cells and lead to T-cell differentiation, favoring Th1 and Th17 cells.<sup>82</sup> Moreover, DCR3 skews



**FIGURE 1** Function of different subtypes of Th cells in PE. Note. TH: T helper; PE: Preeclampsia; Treg: T regulatory; IL: interleukin; TNF- $\alpha$ : tumor necrosis factor-alpha; NK: Natural killer. A, Adenosine has a strong influence on shifting the Th1/Th2 balance toward Th2. B, Transfusion can elevate the risk of PE by increasing IL-6, TNF- $\alpha$ , NK cells, and AT1-AA. C, The interactions between vitamin D and its receptors in the cytoplasm of Treg and Th17 cells can release TGF- $\beta$ , block IL-6 cytokines, and suppress PE development. D, Decoy receptor 3 can bind to TL1A on immune cells and lead to T-cell differentiation, favoring Th1 and Th17 cells, and finally, causing PE

macrophages to the M2 phenotype and regulates dendritic cell differentiation, resulting in a Th1-suppressing effect and Th2 polarization.<sup>83</sup> Transgenic mice who highly express DCR3 show attenuated Th2 differentiation. DCR3 has different roles in decreasing major histocompatibility complex class II expression and dampening T-cell response to alloantigen via interfering with the LIGHT-TR2/HVEM interaction.<sup>84</sup> In pregnancy, DCR3 expression is detected in preeclamptic placentas and is elevated in the excessive inflammatory activities of PE.<sup>85</sup> Ching Yeh et al<sup>86</sup> revealed that the plasma level of DCR3 in PE patients was meaningfully lower than that of the normal pregnant group, suggesting that a potential involvement of DCR3 in normal pregnancy and the reduced levels of DCR3 may be related to immune dysregulation in PE.

## 8 | MicroRNAs AND THEIR FUNCTION IN PE

Similar to short endogenous regulatory RNAs, miRNAs are approximately 22-24 nucleotides long.<sup>87,88</sup> Their main function is to pair

with the 3'-untranslated region of target mRNA and regulate gene translation. Further, miRNAs exert a key role in different physiological and pathological disorders, and the evidence demonstrates that miRNAs are important regulators in placental development and the immune environment.<sup>89-91</sup> Moreover, miRNAs are highly stable in the plasma, serum, and urine, offering the possibility of miRNA to diagnostic or prognostic markers for different illnesses. Dysregulation in the expression of miRNA or abnormal miRNA levels is found in placentas from women who have pregnancy complications such as PE, recurrent miscarriage, and fetal growth restriction, which can affect cell cycle, migration, adhesion, and antiapoptotic survival.<sup>92,93</sup> Differential expression of miRNA has been revealed in PE patients. As reported in previous research, the expression of miR-181a, miR-210, miR-155, miR-182, miR-196, miR-195, and miR-26 increases while that of miR-144 and miR-223 represents a decrease.<sup>93</sup> The modification of miRNAs is critical in the expansion of PE because of their effect on the regulation of immune response and inflammation.<sup>94</sup> In recent years, the association between alterations in miRNA expression and the function of T lymphocytes or the secretion of

different cytokines has been an interesting subject of research. The dysregulation of miRNA-related T lymphocytes is provided in Table 2.

## 8.1 | Dysregulation of miRNAs related to T lymphocytes in PE

### 8.1.1 | miR-106b/miR-326

miR-106b induces cell cycle progression, modulates cell cycle-related proteins, and controls cell proliferation.<sup>95</sup> The upregulation of miR-106b targets and suppresses two pivotal effectors of the TGF- $\beta$  signaling pathway, including cyclin-dependent kinase inhibitor 1A (CDKN1A/p21) and BCL2L1/Bim.<sup>96</sup> TGF- $\beta$  is necessary for the maturation and differentiation of Treg cells, and any dysregulation in this miRNA may alter the activity of Treg cells.<sup>96</sup> Moreover, miR-326 affects the differentiation of Th17 lymphocytes by preventing Ets-1 activity.<sup>97</sup> It has been proven that Ets-1 has an important role in the modulation of Treg and Th17 cell responses.<sup>98</sup> Eghbal-fard et al<sup>46</sup> reported that the expression of miR-106b and miR-326 is upregulated in PE women, and the dysregulation of these two miRNAs can affect the Th17/Treg balance and make a shift toward Th17 response in PE patients.

### 8.1.2 | miR-106\_363 cluster

The cluster of miR-106\_363 is located on chromosome X. A previous study revealed that the upregulation of miR-20a, miR-17, and miR-106a can lead to macrophage activation and increase Th1 cells as compared with Th2 cells.<sup>99</sup> Thus, this miRNA cluster is expressed at the intermediate levels in the Th2, the highest level in the Th1, and the lowest level in Th17 cells.<sup>100</sup> The expression of miR-363-3p, miR-106a, and miR-18b can decrease IL-17 gene expression and prevent the production and activity of IL-17a and Th17, respectively. miR-20b

and miR-182 are the other members of the miR-106\_363 cluster and target ROR $\gamma$ t and the signal transducer and the activator of transcription 3 (STAT3), which are Th17 cell transcription factors and suppress Th17 cells and experimental autoimmune encephalitis.<sup>101</sup> Therefore, the expression of the cluster of miR-106\_363 can impede Th17-mediated inflammations such as chronic disease, asthma, and PE. Therefore, using miR-18b, miR-106a, and miR-363-3p may be a therapeutic approach for Th17 cell-mediated preeclampsia.

### 8.1.3 | miR-210

miR-210 has been implicated in several pathophysiological pathways such as cancer, apoptosis, and oxidative stress.<sup>102,103</sup> Recently, it has been found that miR-210 levels meaningfully increase in the placental and plasma-derived samples from PE patients.<sup>104</sup> Trophoblast cell migration and invasion reduce following miR-210 overexpression in PE.<sup>93</sup> Further, this component causes reductions in STAT6 and IL-4 levels that contribute to the expansion of PE.<sup>105</sup> Additionally, the overexpression of miRNA-210 can suppress the expression of Foxp3 and impair Treg cell functions.<sup>106</sup> The expression of miR-210, which targets Foxp3 and inhibits Treg cell functions, is high in the PE placenta.<sup>107</sup>

### 8.1.4 | miR-155

miR-155 is processed from the human nonprotein coding exon 3 of B-cell integration cluster RNA<sup>108</sup> and is effectively upregulated in T and B cells as a response to antigen stimulation.<sup>109,110</sup> In addition, this miRNA could be induced by Toll-like receptor ligation in dendritic cells and macrophages.<sup>111,112</sup> miR-155 is extremely expressed in Treg cells and is essential for the production of normal thymic-derived Treg cells.<sup>113</sup> Altered miR-155 has been reported in adverse pregnancy outcomes including recurrent miscarriage, recurrent implantation failure, and preeclampsia, suggesting that miR-155 plays

miRNAs	Expression level in PE	Function
miR-106b miR-326	↑	Increasing ROR $\gamma$ t mRNA and elevating Th17 cell differentiation, and decreasing Foxp3 mRNA expression in Treg cells
miR-363-3p miR-106a miR-18b	↑	Increasing IL-17 gene expression and high production of IL-17a and Th17 activity
miR-20b miR-182	↑	High expression of ROR $\gamma$ t and STAT3, which are transcription factors of Th17 cells
miRNA-210	↑	Suppressing expression of Foxp3 and impairing Treg cells function
miR-155	↑	induced podocyte apoptosis through increasing in IL-17 production
miR-320a	↑	Repressing IL-4 expression via binding to 3' untranslated region

**TABLE 2** Dysregulation of miRNAs related to T cell subsets in PE

an important role in modulating the immune system during pregnancy. Furthermore, it is essential for inducing Treg cells regarding achieving tolerance and preventing miscarriage.<sup>113</sup> As reported by one study, in PE following the maternal and fetal inflammation, the overexpression of miR-155 was found in the placenta.<sup>114</sup> Recently, it has been revealed that the upregulation of miR-155 induced podocyte apoptosis through increasing in IL-17 production.<sup>115</sup>

### 8.1.5 | miR-320a

The expression of miR-320a, which is a tumor inhibitor via decreasing the expression of several target genes, is downregulated in various tumor cells.<sup>116</sup> According to previous research, miR-320a has low expression in human non-small lung cancer cells and acts as a crucial regulator in gastric cancer by targeting Ras-related protein Rab-14.<sup>117</sup> Overexpression of miR-320 isoforms in activated T cells can reduce the TGF- $\beta$  signaling pathway by suppressing TGFBR2 and Smad2 genes.<sup>118</sup> miR-320a has an important impact on the invasion and proliferation of trophoblast cells.<sup>119</sup> Recently, it has been found that miR-320a levels increase in serum of PE patients.<sup>120</sup> Xi et al<sup>119</sup> reported that the upregulation of miR-320a in PE patients may contribute to the expansion of PE through repressing IL-4 expression via binding to the 3' untranslated region, which leads to the inhibition of trophoblast proliferation and invasion. The results of this study suggested that the miR-320a/IL-4 pathway may represent a new therapeutic approach for PE.

## 9 | CONCLUSION AND PROSPECT

T lymphocyte cells have four main subsets including Th1, Th2, Th17, and Treg. A normal association between these T cells is necessary for preventing pregnancy disorders such as PE. In recent years, studies have examined the variation of T-cell functions, as well as the cytokines and chemokines that they release in PE. High levels of adenosine and DCR3 or low levels of 25 (OH) vitamin D can lead to imbalances in the Th1/Th2 and Th17/Treg ratios, and eventually, cause PE. Moreover, changing the expression of some miRNAs related to T cells can be a potential molecule for the development of inflammation and PE. In the future, using some blockers for suppressing cytokine or chemokine receptors on T cells or using some siRNAs for inhibiting or activating miRNAs related to T cells can affect the functions of T cells, thus decreasing pregnancy inflammation and preventing PE disease.

### ACKNOWLEDGEMENTS

Authors would like to acknowledge Tabriz University of Medical Sciences, Tabriz, Iran, for their great help.

### DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were created or analyzed in this study.

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### REFERENCES

- Lu H, Hu R. The role of immunity in the pathogenesis and development of pre-eclampsia. *Scand J Immunol.* 2019;90(5):e12756. <https://doi.org/10.1111/sji.12756>
- Brown MA, Magee LA, Kenny LC, et al. Hypertensive disorders of pregnancy: ISSHP classification, diagnosis, and management recommendations for international practice. *Hypertension.* 2018;72(1):24-43. <https://doi.org/10.1161/HYPERTENSIONAHA.117.10803>
- Gill M, Motta-Mejia C, Kandzija N, et al. Placental syncytiotrophoblast-derived extracellular vesicles carry active NEP (Nephrilysin) and are increased in preeclampsia. *Hypertension.* 2019;73(5):1112-1119. <https://doi.org/10.1161/HYPERTENSIONAHA.119.12707>
- Soltani-Zangbar MS, Pahlavani B, Zolghadri J, Ghareesi-Fard B. Angiotensin type 2 receptor gene polymorphisms and susceptibility to preeclampsia. *J Reprod Infertil.* 2018;19(2):95-99.
- Redman CWG. Pre-eclampsia and the placenta. *Placenta.* 1991;12(4):301-308. [https://doi.org/10.1016/0143-4004\(91\)90339-H](https://doi.org/10.1016/0143-4004(91)90339-H)
- Bulmer JN. Human endometrial lymphocytes in normal pregnancy and pregnancy loss. *Ann NY Acad Sci.* 1994;734(1 The Human End):185-192. <https://doi.org/10.1111/j.1749-6632.1994.tb21746.x>
- Danaei S, Ghorbani F, Ahmadi M, et al. IL-10-producing B cells play important role in the pathogenesis of recurrent pregnancy loss. *Int Immunopharmacol.* 2020;87:106806. <https://doi.org/10.1016/j.intimp.2020.106806>
- Wang W, Sung N, Gilman-Sachs A, Kwak-Kim J. T Helper (Th) cell profiles in pregnancy and recurrent pregnancy losses: Th1/Th2/Th9/Th17/Th22/Tfh cells. *Front Immunol.* 2020;11:2025. <https://doi.org/10.3389/fimmu.2020.02025>
- Azizi R, Soltani-Zangbar MS, Sheikhsari G, et al. Metabolic syndrome mediates inflammatory and oxidative stress responses in patients with recurrent pregnancy loss. *J Reprod Immunol.* 2019;133:18-26. <https://doi.org/10.1016/j.jri.2019.05.001>
- Guerin LR, Prins JR, Robertson SA. Regulatory T-cells and immune tolerance in pregnancy: a new target for infertility treatment? *Hum Reprod Update.* 2009;15(5):517-535. <https://doi.org/10.1093/humupd/dmp004>
- Kamrani A, Soltani-Zangbar MS, Shiri S, et al. TIGIT and CD155 as immune-modulator receptor and ligand on CD4 + T cells in preeclampsia patients. *Immunol Invest.* 2021;1-16. <https://doi.org/10.1080/08820139.2021.1904976>
- Sakaguchi S, Miyara M, Costantino CM, Hafler DA. FOXP3 + regulatory T cells in the human immune system. *Nat Rev Immunol.* 2010;10(7):490-500. <https://doi.org/10.1038/nri2785>
- Hosseini A, Dolati S, Hashemi V, Abdollahpour-Alitappeh M, Yousefi M. Regulatory T and T helper 17 cells: their roles in preeclampsia. *J Cell Physiol.* 2018;233(9):6561-6573. <https://doi.org/10.1002/jcp.26604>
- Thornton AM, Korty PE, Tran DQ, et al. Expression of helios, an Ikaros transcription factor family member, differentiates thymic-derived from peripherally induced Foxp3 + T regulatory cells. *J Immunol.* 2010;184(7):3433-3441. <https://doi.org/10.4049/jimmunol.0904028>
- Rowe JH, Ertelt JM, Xin L, Way SS. Pregnancy imprints regulatory memory that sustains anergy to fetal antigen. *Nature.* 2012;490(7418):102-106. <https://doi.org/10.1038/nature11462>
- Pongcharoen S, Supalap K. Short communication: interleukin-17 increased progesterone secretion by JEG-3 human choriocarcinoma

- cells. *Am J Reprod Immunol*. 2009;61(4):261-264. <https://doi.org/10.1111/j.1600-0897.2009.00693.x>
17. Nakashima A, Ito M, Yoneda S, Shiozaki A, Hidaka T, Saito S. Short communication: circulating and decidual Th17 cell levels in healthy pregnancy. *Am J Reprod Immunol*. 2010;63(2):104-109. <https://doi.org/10.1111/J.1600-0897.2009.00771.X>
  18. Yousefzadeh Y, Soltani-Zangbar MS, Hemmatzadeh M, et al. Fetomaternal immune tolerance: crucial mechanisms of tolerance for successful pregnancy in humans. *Immunol Invest*. 2021;1-18. <https://doi.org/10.1080/08820139.2021.1909061>
  19. Liu F, Guo J, Tian T, et al. Placental trophoblasts shifted Th1/Th2 balance toward Th2 and inhibited Th17 immunity at fetomaternal interface. *APMIS*. 2011;119(9):597-604. <https://doi.org/10.1111/j.1600-0463.2011.02774.x>
  20. Harmon AC, Cornelius DC, Amaral LM, et al. The role of inflammation in the pathology of preeclampsia. *Clin Sci*. 2016;130(6):409-419. <https://doi.org/10.1042/CS20150702>
  21. Arriaga-Pizano L, Jimenez-Zamudio L, Vadillo-Ortega F, Martinez-Flores A, Herrerias-Canedo T, Hernandez-Guerrero C. The predominant Th1 cytokine profile in maternal plasma of preeclamptic women is not reflected in the chorionic and fetal compartments. *J Soc Gynecol Investig*. 2005;12(5):335-342. <https://doi.org/10.1016/j.jsg.2005.02.005>
  22. Borzychowski AM, Croy BA, Chan WL, Redman CWG, Sargent IL. Changes in systemic type 1 and type 2 immunity in normal pregnancy and pre-eclampsia may be mediated by natural killer cells. *Eur J Immunol*. 2005;35(10):3054-3063. <https://doi.org/10.1002/eji.200425929>
  23. Miko E, Szereday L, Barakonyi A, Jarkovich A, Varga P, Szekeres-Bartho J. Immunoactivation in preeclampsia: V $\delta$ 2+ and regulatory T cells during the inflammatory stage of disease. *J Reprod Immunol*. 2009;80(1-2):100-108. <https://doi.org/10.1016/j.jri.2009.01.003>
  24. Hashemi V, Dolati S, Hosseini A, Gharibi T, Danaii S, Yousefi M. Natural killer T cells in preeclampsia: an updated review. *Biomed Pharmacother*. 2017;95:412-418. <https://doi.org/10.1016/j.biopha.2017.08.077>
  25. Jianjun Z, Yali H, Zhiqun W, Mingming Z, Xia Z. Original article: imbalance of T-cell transcription factors contributes to the Th1 type immunity predominant in pre-eclampsia. *Am J Reprod Immunol*. 2009;63(1):38-45. <https://doi.org/10.1111/j.1600-0897.2009.00763.x>
  26. Ribeiro VR, Romao-Veiga M, Romagnoli GG, et al. Association between cytokine profile and transcription factors produced by T-cell subsets in early- and late-onset pre-eclampsia. *Immunology*. 2017;152(1):163-173. <https://doi.org/10.1111/imm.12757>
  27. Robertson SA, Green ES, Care AS, et al. Therapeutic potential of regulatory T cells in preeclampsia-opportunities and challenges. *Front Immunol*. 2019;10:478. <https://doi.org/10.3389/fimmu.2019.00478>
  28. Darmochwal-Kolarz D, Kludka-Sternik M, Tabarkiewicz J, et al. The predominance of Th17 lymphocytes and decreased number and function of Treg cells in preeclampsia. *J Reprod Immunol*. 2012;93(2):75-81. <https://doi.org/10.1016/j.jri.2012.01.006>
  29. Toldi G, Vászrhelyi ZE, Rigó J, et al. Prevalence of regulatory T-cell subtypes in preeclampsia. *Am J Reprod Immunol*. 2015;74(2):110-115. <https://doi.org/10.1111/aji.12380>
  30. Feger U, Tolosa E, Huang YH, et al. HLA-G expression defines a novel regulatory T-cell subset present in human peripheral blood and sites of inflammation. *Blood*. 2007;110(2):568-577. <https://doi.org/10.1182/blood-2006-11-057125>
  31. Pankratz S, Bittner S, Herrmann AM, et al. Human CD4<sup>+</sup> HLA-G<sup>+</sup> regulatory T cells are potent suppressors of graft-versus-host disease in vivo. *FASEB J*. 2014;28(8):3435-3445. <https://doi.org/10.1096/fj.14-251074>
  32. Zare M, Namavar Jahromi B, Ghahesi-Fard B. Analysis of the frequencies and functions of CD4+CD25+CD127low/neg, CD4+HLA-G+, and CD8+HLA-G+ regulatory T cells in pre-eclampsia. *J Reprod Immunol*. 2019;133:43-51. <https://doi.org/10.1016/j.jri.2019.06.002>
  33. Saito S. Th17 cells and regulatory T cells: new light on pathophysiology of preeclampsia. *Immunol Cell Biol*. 2010;88(6):615-617. <https://doi.org/10.1038/icb.2010.68>
  34. Daraei N, Ghafourian M, Ghadiri A, Amari A, Najafian M, Rokhafrooz S. Evaluation of exhausted regulatory T cells in preeclampsia. *Iran J Immunol*. 2019;16(2):163-169. <https://doi.org/10.22034/iji.2019.80259>
  35. Tian M, Zhang Y, Liu Z, Sun G, Mor G, Liao A. The PD-1/PD-L1 inhibitory pathway is altered in pre-eclampsia and regulates T cell responses in pre-eclamptic rats. *Sci Rep*. 2016;6(1):1-14. <https://doi.org/10.1038/srep27683>
  36. Jafarzadeh S, Ahmadi M, Dolati S, et al. Intravenous immunoglobulin G treatment increases live birth rate in women with recurrent miscarriage and modulates regulatory and exhausted regulatory T cells frequency and function. *J Cell Biochem*. 2019;120(4):5424-5434. <https://doi.org/10.1002/jcb.27821>
  37. Sharpe AH, Wherry EJ, Ahmed R, Freeman GJ. The function of programmed cell death 1 and its ligands in regulating autoimmunity and infection. *Nat Immunol*. 2007;8(3):239-245. <https://doi.org/10.1038/ni1443>
  38. Zhang Y-H, Tian M, Tang M-X, Liu Z-Z, Liao A-H. Recent insight into the role of the PD-1/PD-L1 pathway in fetomaternal tolerance and pregnancy. *Am J Reprod Immunol*. 2015;74(3):201-208. <https://doi.org/10.1111/aji.12365>
  39. Zhang Y, Liu Z, Tian M, et al. The altered PD-1/PD-L1 pathway delivers the 'one-two punch' effects to promote the Treg/Th17 imbalance in pre-eclampsia. *Cell Mol Immunol*. 2018;15(7):710-723. <https://doi.org/10.1038/cmi.2017.70>
  40. Saito S, Nakashima A, Shima T, Ito M. Th1/Th2/Th17 and regulatory T-cell paradigm in pregnancy. *Am J Reprod Immunol*. 2010;63(6):601-610. <https://doi.org/10.1111/j.1600-0897.2010.00852.x>
  41. Anvari F, Dabagh-Gorjani F, Soltani-Zangbar M-S, Kamali-Sarvestani E, Malek-Hosseini Z, Ghahesi-Fard B. Investigating the association of IL-17A and IL-17F with susceptibility to preeclampsia in Iranian women. Vol 12. Shiraz Institute for Cancer Research; 2015. Accessed March 22, 2021. [https://iji.sums.ac.ir/article\\_16741.html](https://iji.sums.ac.ir/article_16741.html)
  42. Wallace K, Richards S, Dhillon P, et al. CD4<sup>+</sup> T-helper cells stimulated in response to placental ischemia mediate hypertension during pregnancy. *Hypertension*. 2011;57(5):949-955. <https://doi.org/10.1161/HYPERTENSIONAHA.110.168344>
  43. Parrish MR, Murphy SR, Rutland S, et al. The effect of immune factors, tumor necrosis factor-, and agonistic autoantibodies to the angiotensin II type I receptor on soluble fms-like tyrosine-1 and soluble endoglin production in response to hypertension during pregnancy. *Am J Hypertens*. 2010;23(8):911-916. <https://doi.org/10.1038/ajh.2010.70>
  44. Cornelius DC, Amaral LM, Wallace K, et al. Reduced uterine perfusion pressure T-helper 17 cells cause pathophysiology associated with preeclampsia during pregnancy. *Am J Physiol Regul Integr Comp Physiol*. 2016;311(6):R1192-R1199. <https://doi.org/10.1152/ajpregu.00117.2016>
  45. Travis OK, White D, Pierce WA, et al. Chronic infusion of interleukin-17 promotes hypertension, activation of cytolytic natural killer cells, and vascular dysfunction in pregnant rats. *Physiol Rep*. 2019;7(7):e14038. <https://doi.org/10.14814/phy2.14038>
  46. Eghbal-Fard S, Yousefi M, Heydarlou H, et al. The imbalance of Th17/Treg axis involved in the pathogenesis of preeclampsia. *J Cell Physiol*. 2019;234(4):5106-5116. <https://doi.org/10.1002/jcp.27315>
  47. Miko E, Szereday L, Barakonyi A, Jarkovich A, Varga P, Szekeres-Bartho J. Original article: the role of invariant NKT cells in



- pre-eclampsia. *Am J Reprod Immunol.* 2008;60(2):118-126. <https://doi.org/10.1111/j.1600-0897.2008.00603.x>
48. Barakonyi A, Miko E, Szereday L, et al. Cell death mechanisms and potentially cytotoxic natural immune cells in human pregnancies complicated by preeclampsia. *Reprod Sci.* 2014;21(2):155-166. <https://doi.org/10.1177/1933719113497288>
  49. Nancy P, Tagliani E, Tay CS, Asp P, Levy DE, Erlebacher A. Chemokine gene silencing in decidual stromal cells limits T cell access to the maternal-fetal interface. *Science.* 2012;336(6086):1317-1321. <https://doi.org/10.1126/science.1220030>
  50. Di Castro MA, Trettel F, Milior G, Maggi L, Ragozzino D, Limatola C. The chemokine CXCL16 modulates neurotransmitter release in hippocampal CA1 area. *Sci Rep.* 2016;6(1):1-13. <https://doi.org/10.1038/srep34633>
  51. Izquierdo MC, Martin-Cleary C, Fernandez-Fernandez B, et al. CXCL16 in kidney and cardiovascular injury. *Cytokine Growth Factor Rev.* 2014;25(3):317-325. <https://doi.org/10.1016/j.cytogfr.2014.04.002>
  52. Sheikine Y, Sirsjo A. CXCL16/SR-PSOX-A friend or a foe in atherosclerosis? *Atherosclerosis.* 2008;197(2):487-495. <https://doi.org/10.1016/j.atherosclerosis.2007.11.034>
  53. Stone M, Hayward J, Huang C, E. Huma Z, Sanchez J. Mechanisms of regulation of the chemokine-receptor network. *Int J Mol Sci.* 2017;18(2):342. <https://doi.org/10.3390/ijms18020342>
  54. Tok A, Seyithanoğlu M, Ozer A, Er kayıran U, Karaküçük S, Çelebi A. The serum level of soluble CXCL16 is increased in preeclampsia and associated with hepatic/renal damage. *J Matern Fetal Neonatal Med.* 2019;34(9):1435-1440. <https://doi.org/10.1080/14767058.2019.1638361>
  55. Ivashkiv LB. IFN $\gamma$ : signalling, epigenetics and roles in immunity, metabolism, disease and cancer immunotherapy. *Nat Rev Immunol.* 2018;18(9):545-558. <https://doi.org/10.1038/s41577-018-0029-z>
  56. Yang Y, Su X, Xu W, Zhou R. Interleukin-18 and interferon gamma levels in preeclampsia: a systematic review and meta-analysis. *Am J Reprod Immunol.* 2014;72(5):504-514. <https://doi.org/10.1111/aji.12298>
  57. Tokunaga R, Zhang W, Naseem M, et al. CXCL9, CXCL10, CXCL11/CXCR3 axis for immune activation – a target for novel cancer therapy. *Cancer Treat Rev.* 2018;63:40-47. <https://doi.org/10.1016/j.ctrv.2017.11.007>
  58. Noroozi Karimabad M, Khanamani Falahati-Pour S, Hassanshahi G. Significant role(s) of CXCL12 and the SDF-1 3'a genetic variant in the pathogenesis of multiple sclerosis. *NeuroImmunoModulation.* 2017;23(4):197-208. <https://doi.org/10.1159/000449427>
  59. Moreno-Eutimio MA, Tovar-Rodríguez JM, Vargas-Avila K, et al. Increased serum levels of inflammatory mediators and low frequency of regulatory T cells in the peripheral blood of preeclamptic Mexican women. *BioMed Res Int.* 2014;2014:1-8. <https://doi.org/10.1155/2014/413249>
  60. Darakhshan S, Hassanshahi G, Mofidifar Z, Soltani B, Karimabad MN. CXCL9/CXCL10 angiostasis CXC-chemokines in parallel with the CXCL12 as an angiogenesis CXC-chemokine are variously expressed in pre-eclamptic-women and their neonates. *Pregnancy Hypertens.* 2019;17:36-42. <https://doi.org/10.1016/j.preghy.2019.05.001>
  61. Whitelaw E. Epigenetics: sins of the fathers, and their fathers. *Eur J Hum Genet.* 2006;14(2):131-132. <https://doi.org/10.1038/sj.ejhg.5201567>
  62. Drake AJ, Walker BR. The intergenerational effects of fetal programming: non-genomic mechanisms for the inheritance of low birth weight and cardiovascular risk. *J Endocrinol.* 2004;180(1):1-16. <https://doi.org/10.1677/joe.0.1800001>
  63. Ji J-L, Muyayalo KP, Zhang Y-H, Hu X-H, Liao A-H. Immunological function of vitamin D during human pregnancy. *Am J Reprod Immunol.* 2017;78(2):e12716. <https://doi.org/10.1111/aji.12716>
  64. Washington K, Ghosh S, Reeves IV. A review: molecular concepts and common pathways involving vitamin D in the pathophysiology of preeclampsia. *Open J Obstet Gynecol.* 2018;08(03):198-229. <https://doi.org/10.4236/ojog.2018.83023>
  65. Song J, Li Y, An R. Vitamin D restores angiogenic balance and decreases tumor necrosis factor- $\alpha$  in a rat model of pre-eclampsia. *J Obstet Gynaecol Res.* 2017;43(1):42-49. <https://doi.org/10.1111/jog.13186>
  66. Muyayalo KP, Huang X, Qian Z, Li Z, Mor G, Liao A. Low circulating levels of vitamin D may contribute to the occurrence of preeclampsia through deregulation of Treg /Th17 cell ratio. *Am J Reprod Immunol.* 2019;82(4):e13168. <https://doi.org/10.1111/aji.13168>
  67. Zhou Q, Qin S, Zhang J, Zhon L, Pen Z, Xing T. 1,25(OH)2D3 induces regulatory T cell differentiation by influencing the VDR/PLC- $\gamma$ 1/TGF- $\beta$ 1/pathway. *Mol Immunol.* 2017;91:156-164. <https://doi.org/10.1016/j.molimm.2017.09.006>
  68. Nonn L, Peng L, Feldman D, Peehl DM. Inhibition of p38 by vitamin D reduces interleukin-6 production in normal prostate cells via mitogen-activated protein kinase phosphatase 5: implications for prostate cancer prevention by vitamin D. *Cancer Res.* 2006;66(8):4516-4524. <https://doi.org/10.1158/0008-5472.CAN-05-3796>
  69. Xiao J-P, Lv J-X, Yin Y-X, et al. Lower maternal and fetal vitamin D status and higher placental and umbilical vitamin D receptor expression in preeclamptic pregnancies. *Int J Clin Exp Pathol.* 2017;10(11):10841-10851. Accessed January 29, 2021. <http://www.ncbi.nlm.nih.gov/pubmed/31966427>
  70. Serrano NC, Guío E, Quintero-Lesmes DC, et al. Vitamin D deficiency and pre-eclampsia in Colombia: PREVITd study. *Pregnancy Hypertens.* 2018;14:240-244. <https://doi.org/10.1016/j.preghy.2018.03.006>
  71. Sahu M, Tripathy S, Bhuyan P. Association of maternal serum vitamin D level with preeclampsia or eclampsia and its relationship with neonatal outcome and neonatal serum calcium level. *Int J Reprod Contracept Obstet Gynecol.* 2017;6(12):5580. <https://doi.org/10.18203/2320-1770.ijrcog20175285>
  72. Bodnar LM, Simhan HN, Catov JM, et al. Maternal vitamin D status and the risk of mild and severe preeclampsia. *Epidemiology.* 2014;25(2):207-214. <https://doi.org/10.1097/EDE.0000000000000039>
  73. Zhao X, Fang R, Yu R, Chen D, Zhao J, Xiao J. Maternal vitamin D status in the late second trimester and the risk of severe preeclampsia in Southeastern China. *Nutrients.* 2017;9(2):138. <https://doi.org/10.3390/nu9020138>
  74. Robinson CJ, Alanis MC, Wagner CL, Hollis BW, Johnson DD. Plasma 25-hydroxyvitamin D levels in early-onset severe preeclampsia. *Am J Obstet Gynecol.* 2010;203(4):366.e1-366.e6. <https://doi.org/10.1016/j.ajog.2010.06.036>
  75. Zidek Z. Adenosine - cyclic AMP pathways and cytokine expression. *Eur Cytokine Netw.* 1999;10(3):319-328. Accessed February 1, 2021. [http://www.jle.com/fr/revues/ecn/e-docs/adenosine\\_cyclic\\_amp\\_pathways\\_and\\_cytokine\\_expression\\_90308/article.phtml?tab=texte](http://www.jle.com/fr/revues/ecn/e-docs/adenosine_cyclic_amp_pathways_and_cytokine_expression_90308/article.phtml?tab=texte)
  76. Elenkov IJ, Chrousos GP, Wilder RL. Neuroendocrine regulation of IL-12 and TNF- $\alpha$ /IL-10 balance: clinical implications. *Ann NY Acad Sci.* 2006;917(1):94-105. <https://doi.org/10.1111/j.1749-6632.2000.tb05374.x>
  77. Yoneyama Y, Suzuki S, Sawa R, Kiyokawa Y, Power GG, Araki T. Plasma adenosine levels and P-selectin expression on platelets in preeclampsia. *Obstet Gynecol.* 2001;97(3):366-370. [https://doi.org/10.1016/S0029-7844\(00\)01184-4](https://doi.org/10.1016/S0029-7844(00)01184-4)
  78. Yoneyama Y, Suzuki S, Sawa R, Yoneyama K, Power GG, Araki T. Relation between adenosine and T-helper 1/T-helper 2 imbalance in women with preeclampsia. *Obstet Gynecol.* 2002;99(4):641-646. [https://doi.org/10.1016/S0029-7844\(02\)01657-5](https://doi.org/10.1016/S0029-7844(02)01657-5)

79. Lin WW, Hsieh SL. Decoy receptor 3: a pleiotropic immunomodulator and biomarker for inflammatory diseases, autoimmune diseases and cancer. *Biochem Pharmacol.* 2011;81(7):838-847. <https://doi.org/10.1016/j.bcp.2011.01.011>
80. Li TH, Liu CW, Lee PC, et al. Decoy receptor 3 analogous supplement protects steatotic rat liver from ischemia-reperfusion injury. *J Chin Med Assoc.* 2017;80(7):391-400. <https://doi.org/10.1016/j.jcma.2016.11.008>
81. Pitti RM, Marsters SA, Lawrence DA, et al. Genomic amplification of a decoy receptor for fas ligand in lung and colon cancer. *Nature.* 1998;396(6712):699-703. <https://doi.org/10.1038/25387>
82. Migone TS, Zhang J, Luo X, et al. TL1A is a TNF-like ligand for DR3 and TR6/DcR3 and functions as a T cell costimulator. *Immunity.* 2002;16(3):479-492. [https://doi.org/10.1016/S1074-7613\(02\)00283-2](https://doi.org/10.1016/S1074-7613(02)00283-2)
83. Hsieh SL, Lin WW. Decoy receptor 3: an endogenous immunomodulator in cancer growth and inflammatory reactions. *J Biomed Sci.* 2017;24(1):39. <https://doi.org/10.1186/s12929-017-0347-7>
84. Zhang J, Salcedo TW, Wan X, et al. Modulation of T-cell responses to alloantigens by TR6/DcR3. *J Clin Invest.* 2001;107(11):1459-1468. <https://doi.org/10.1172/JCI12159>
85. Chang YC, Chen TC, Lee CT, et al. Epigenetic control of MHC class II expression in tumor-associated macrophages by decoy receptor 3. *Blood.* 2008;111(10):5054-5063. <https://doi.org/10.1182/blood-2007-12-130609>
86. Yeh CC, Yang MJ, Lussier EC, et al. Low plasma levels of decoy receptor 3 (DcR3) in the third trimester of pregnancy with preeclampsia. *Taiwan J Obstet Gynecol.* 2019;58(3):349-353. <https://doi.org/10.1016/j.tjog.2019.03.011>
87. Bartel DP. MicroRNAs: target recognition and regulatory functions. *Cell.* 2009;136(2):215-233. <https://doi.org/10.1016/j.cell.2009.01.002>
88. Kamrani A, Alipourfard I, Ahmadi-Khiavi H, et al. The role of epigenetic changes in preeclampsia. *BioFactors.* 2019;45(5):712-724. <https://doi.org/10.1002/biof.1542>
89. Shomali N, Hemmatzadeh M, Yousefzadeh Y, et al. Exosomes: emerging biomarkers and targets in folliculogenesis and endometriosis. *J Reprod Immunol.* 2020;142:103181. <https://doi.org/10.1016/j.jri.2020.103181>
90. Wienholds E, Plasterk RHA. MicroRNA function in animal development. *FEBS Lett.* 2005;579(26):5911-5922. <https://doi.org/10.1016/j.febslet.2005.07.070>
91. Bidarimath M, Khalaj K, Wessels JM, Tayade C. MicroRNAs, immune cells and pregnancy. *Cell Mol Immunol.* 2014;11(6):538-547. <https://doi.org/10.1038/cmi.2014.45>
92. Pineles BL, Romero R, Montenegro D, et al. Distinct subsets of microRNAs are expressed differentially in the human placentas of patients with preeclampsia. *Am J Obstet Gynecol.* 2007;196(3):261.e1-261.e6. <https://doi.org/10.1016/j.ajog.2007.01.008>
93. Hemmatzadeh M, Shomali N, Yousefzadeh Y, Mohammadi H, Ghasemzadeh A, Yousefi M. MicroRNAs: small molecules with a large impact on pre-eclampsia. *J Cell Physiol.* 2020;235(4):3235-3248. <https://doi.org/10.1002/jcp.29286>
94. Wang Z, Wang P, Wang Z, et al. MiRNA-548c-5p downregulates inflammatory response in preeclampsia via targeting PTPRO. *J Cell Physiol.* 2019;234(7):11149-11155. <https://doi.org/10.1002/jcp.27758>
95. Zhang Z, Liu H, Shi Y, et al. Increased circulating Th22 cells correlated with Th17 cells in patients with severe preeclampsia. *Hypertens Pregnancy.* 2017;36(1):100-107. <https://doi.org/10.1080/10641955.2016.1239737>
96. Esmailzadeh S, Mansoori B, Mohammadi A, Baradaran B. Regulatory roles of micro-RNAs in T cell autoimmunity. *Immunol Invest.* 2017;46(8):864-879. <https://doi.org/10.1080/08820139.2017.1373901>
97. Zolfaghari MA, Motavalli R, Soltani-Zangbar MS, et al. A new approach to the preeclampsia puzzle; microRNA-326 in CD4+ lymphocytes might be as a potential suspect. *J Reprod Immunol.* 2021;145:103317. <https://doi.org/10.1016/j.jri.2021.103317>
98. Du C, Liu C, Kang J, et al. MicroRNA miR-326 regulates TH-17 differentiation and is associated with the pathogenesis of multiple sclerosis. *Nat Immunol.* 2009;10(12):1252-1259. <https://doi.org/10.1038/ni.1798>
99. Sasaki K, Kohanbash G, Hoji A, et al. MiR-17-92 expression in differentiated T cells - implications for cancer immunotherapy. *J Transl Med.* 2010;8(1):1-12. <https://doi.org/10.1186/1479-5876-8-17>
100. Kästle M, Bartel S, Geillinger-Kästle K, et al. microRNA cluster 106a-363 is involved in T helper 17 cell differentiation. *Immunology.* 2017;152(3):402-413. <https://doi.org/10.1111/imm.12775>
101. Ingwersen J, Menge T, Wingerath B, et al. Natalizumab restores aberrant miRNA expression profile in multiple sclerosis and reveals a critical role for miR-20b. *Ann Clin Transl Neurol.* 2015;2(1):43-55. <https://doi.org/10.1002/acn3.152>
102. Muralimanoharan S, Guo C, Myatt L, Maloyan A. Sexual dimorphism in MIR-210 expression and mitochondrial dysfunction in the placenta with maternal obesity. *Int J Obes.* 2015;39(8):1274-1281. <https://doi.org/10.1038/ijo.2015.45>
103. Gee HE, Camps C, Buffa FM, et al. hsa-miR-210 is a marker of tumor hypoxia and a prognostic factor in head and neck cancer. *Cancer.* 2010;116(9):2148-2158. <https://doi.org/10.1002/cncr.25009>
104. Lv Y, Lu C, Ji X, et al. Roles of microRNAs in preeclampsia. *J Cell Physiol.* 2019;234(2):1052-1061. <https://doi.org/10.1002/jcp.27291>
105. Kopriva SE, Chiasson VL, Mitchell BM, Chatterjee P. TLR3-induced placental miR-210 down-regulates the STAT6/interleukin-4 pathway. *PLoS One.* 2013;8(7):67760. <https://doi.org/10.1371/journal.pone.0067760>
106. Zhao M, Wang LT, tao, Liang G-P, et al. Up-regulation of microRNA-210 induces immune dysfunction via targeting FOXP3 in CD4+ T cells of psoriasis vulgaris. *Clin Immunol.* 2014;150(1):22-30. <https://doi.org/10.1016/j.clim.2013.10.009>
107. Chen J, Zhao L, Wang D, et al. Contribution of regulatory T cells to immune tolerance and association of microRNA-210 and Foxp3 in preeclampsia. *Mol Med Rep.* 2019;19(2):1150-1158. <https://doi.org/10.3892/mmr.2018.9733>
108. Elton TS, Selemo H, Elton SM, Parinandi NL. Regulation of the MIR155 host gene in physiological and pathological processes. *Gene.* 2013;532(1):1-12. <https://doi.org/10.1016/j.gene.2012.12.009>
109. Thai TH, Calado DP, Casola S, et al. Regulation of the germinal center response by MicroRNA-155. *Science.* 2007;316(5824):604-608. <https://doi.org/10.1126/science.1141229>
110. Grigoryev YA, Kurian SM, Hart T, et al. MicroRNA regulation of molecular networks mapped by global MicroRNA, mRNA, and protein expression in activated T lymphocytes. *J Immunol.* 2011;187(5):2233-2243. <https://doi.org/10.4049/jimmunol.1101233>
111. O'Connell RM, Zhao JL, Rao DS. MicroRNA function in myeloid biology. *Blood.* 2011;118(11):2960-2969. <https://doi.org/10.1182/blood-2011-03-291971>
112. Rodriguez A, Vigorito E, Clare S, et al. Requirement of bic/microRNA-155 for normal immune function. *Science.* 2007;316(5824):608. <https://doi.org/10.1126/SCIENCE.1139253>
113. Schjenken JE, Moldenhauer LM, Zhang B, et al. MicroRNA miR-155 is required for expansion of regulatory T cells to mediate robust pregnancy tolerance in mice. *Mucosal Immunol.* 2020;13(4):609-625. <https://doi.org/10.1038/s41385-020-0255-0>
114. Bounds KR, Chiasson VL, Pan LJ, Gupta S, Chatterjee P. MicroRNAs: new players in the pathobiology of preeclampsia. *Front Cardiovasc Med.* 2017;4:25. <https://doi.org/10.3389/fcvm.2017.00060>
115. Yang X, Zhang J, Ding Y. Association of microRNA-155, interleukin 17A, and proteinuria in preeclampsia. *Medicine.* 2017;96(18):e6509. <https://doi.org/10.1097/MD.0000000000006509>

116. Sun L, Liu B, Lin Z, et al. MiR-320a acts as a prognostic factor and Inhibits metastasis of salivary adenoid cystic carcinoma by targeting ITGB3. *Mol Cancer*. 2015;14(1):96. <https://doi.org/10.1186/s12943-015-0344-y>
117. Li Y, Liu H, Shao J, Xing G. MiR-320a serves as a negative regulator in the progression of gastric cancer by targeting RAB14. *Mol Med Rep*. 2017;16(3):2652-2658. <https://doi.org/10.3892/mmr.2017.6937>
118. Talebi F, Ghourbani S, Voigani M, Noorbakhsh F. miR-320 and inflammation regulation in experimental autoimmune encephalomyelitis through interference with tumor growth factor- $\beta$  signaling pathway. *Immunoregulation*. 2019;2(2):229-238. <https://doi.org/10.32598/immunoregulation.1.4.229>
119. Xie N, Jia Z, Li L. MIR-320a upregulation contributes to the development of preeclampsia by inhibiting the growth and invasion of trophoblast cells by targeting interleukin 4. *Mol Med Rep*. 2019;20(4):3256-3264. <https://doi.org/10.3892/mmr.2019.10574>
120. Zhong Y, Zhu F, Ding Y. Differential microRNA expression profile in the plasma of preeclampsia and normal pregnancies. *Exp Ther Med*. 2019;18(1):826-832. <https://doi.org/10.3892/etm.2019.7637>

**How to cite this article:** Zolfaghari MA, Arefnezhad R, Parhizkar F, et al. T lymphocytes and preeclampsia: The potential role of T-cell subsets and related MicroRNAs in the pathogenesis of preeclampsia. *Am J Reprod Immunol*. 2021;86:e13475. <https://doi.org/10.1111/aji.13475>